



## Increasing the photocatalytic and fungicide activities of $\text{Ag}_3\text{PO}_4$ microcrystals under visible-light irradiation

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### ABSTRACT

The present study reports for the first time the performance of silver phosphate ( $\text{Ag}_3\text{PO}_4$ ) microcrystals as photocatalyst (degradation of Rhodamine B-RhB) and antifungal agent (against *Candida albicans*-*C. albicans*) under visible-light irradiation (455 nm).  $\text{Ag}_3\text{PO}_4$  microcrystals were synthesized by a simple co-precipitation (CP) method at room temperature. The structural and electronic properties of the as-synthesized  $\text{Ag}_3\text{PO}_4$  have been investigated before and after 4 cycles of RhB degradation under visible light using X-ray diffraction (XRD), micro-Raman spectroscopy, UV-Vis spectrophotometer and field emission scanning electron microscopy (FE-SEM) images. The antifungal activity was analyzed in planktonic cells and 48h-biofilm of *C. albicans* by colony forming units (CFU) counting, confocal laser and FE-SE microscopies. Statistical analysis was carried out using SPSS software. Morphological and structural modifications of  $\text{Ag}_3\text{PO}_4$  were observed upon recycling. After 4 recycles, the material maintained its photodegradation property; an eightfold increase in the efficiency of  $\text{Ag}_3\text{PO}_4$  was observed in planktonic cells and a two fold increase in biofilm when irradiated under visible light. Thus, higher antifungal effectiveness against *C. albicans* was obtained when associated with visible-light irradiation.

### 1. Introduction

$\text{Ag}_3\text{PO}_4$  microcrystals have excellent optical properties; photo-luminescent, photocatalytic, bacteriostatic activity; and biocompatibility [1–12], which are important characteristics for materials used in the biomedical field. Moreover,  $\text{Ag}_3\text{PO}_4$  microcrystals have photodegradation potential when exposed to light spectra. In addition, compared to organic photosensitizers,  $\text{Ag}_3\text{PO}_4$  presents advantageous due to their greater stability under different biological conditions, such as pH, ionic strength, pressure, and stability to light [13,14].

*C. albicans* is a commensal fungus that lives in harmony with other microorganisms, but local or systemic factors (HIV, cancer, malnutrition, diabetes, chemotherapy, or radiotherapy) can alter this balance and promote *C. albicans* pathogenicity [15,16]. Denture stomatitis, for example, affects a large number of people using removable oral prostheses. It exhibits multifactorial etiology, including a higher incidence in

women and smokers, inadequate prosthesis use, and microbial colonization, mainly by *Candida* spp [17], and especially by *C. albicans* [18]. The treatment currently used consists of antifungals such as miconazole [19], nystatin, or fluconazole. However, the infection caused by *C. albicans* presents frequent recurrence due to its multifactorial etiology and the lack of antifungal drug efficacy [20].

Among the possibilities for microbial control, the use of microcrystals of semiconductors composed of silver (Ag) has been widely investigated by our research group due to its high activity against bacteria, fungi, and viruses [21–27]. The great advantage of using microcrystals is the non-penetrability in mammalian cells, such as fibroblasts and keratinocytes, due to their larger dimensions [21,28,29], which increase biocompatibility.

$\text{Ag}_3\text{PO}_4$  exhibits photocatalytic activity in the presence of light [11], especially UV light because it presents greater light absorption. For the first time, in the present study, we investigate  $\text{Ag}_3\text{PO}_4$  microcrystals as

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photocatalyst for the degradation of Rhodamine B (RhB) and as biocide against *C. albicans*, in the absence and presence of irradiation in the visible light range (blue) called L- and L+, respectively. This wavelength range was selected because it is commonly employed for the treatment of skin tumors [30], the analysis of melanoma growth rate [31], the treatment of colorectal cancer by the production of reactive oxygen species (ROS) [32], and in dentistry for chemical activation by light-curing of resin-based restorative materials [33,34].

Thus, the aim of this study was to evaluate the photocatalytic and antifungal activity of  $\text{Ag}_3\text{PO}_4$  microcrystals against *C. albicans* and the effect of the presence or absence of visible-light irradiation.

## 2. Results and discussion

### 2.1. Characterization aspects

The analysis of the  $\text{Ag}_3\text{PO}_4$  absorption spectrum (Fig. S1, in the Supporting Information) displays a maximum absorbance value of 1.493 a.u. at the wavelength of 325 nm. Although the absorption peaks in the ultraviolet wavelength (harmful to health) [35–37], the particles were also stimulated by blue light, with an absorbance of 1.252 a.u. at the 455-nm wavelength, because the photon energy from the light used is larger than the value of Egap for  $\text{Ag}_3\text{PO}_4$  (2.24 eV) (Fig. S2). Besides, the 455 nm wavelength used in this research is already used in dental treatments; thus this is an area of interest for our research group for future development of the prevention or treatment of *C. albicans*-related diseases in the oral cavity. Thus, the light used in dental offices covers the visible spectrum of visible light at a specific wavelength of the blue region [38]. Fig. 1 shows the photocatalytic activity of  $\text{Ag}_3\text{PO}_4$  against RhB upon four exposures to blue light (455 nm). RhB shows maximum absorbance between 500- and 600-nm wavelengths, and the degradation of the RhB chromophore was analyzed through the decay of this

absorbance. This decay occurs due to the de-ethylation of the *N,N'*-diethylammonium group in the xanthene ring (chromophore of the RhB) [39]. From the first to the third cycle, there was an increase in the degradation rate of the RhB. In the fourth cycle, the degradation rate decreased. This result can be attributed to the structural and electronic modifications that the material undergoes in the photocorrosion processes for the degradation of the RhB.

The photodegradation results for 455 nm irradiation are shown in Fig. 2A. The RhB concentration ( $C_n/C_0$ ) is presented as function of time, where  $C_n$  is the concentration at time  $t$  and  $C_0$  is the initial concentration. Due to the low concentration of dye and catalyst in the tests, most heterogeneous photocatalytic mechanisms performed by semiconductors are considered pseudo-first-order [40]. Complete photodegradation of RhB after 30 min under visible-light irradiation was reported previously, with  $\text{Ag}_3\text{PO}_4$  synthesized by a facile ion-exchange route [41]. In our study, after 12 min under visible-light irradiation, almost complete photocatalysis is observed in the first and third cycles. This RhB photodegradation time is in agreement with results reported in the literature with  $\text{Ag}_3\text{PO}_4$  nanorods [42]. After 20 min, the rate of degradation was 100%, with the exception of the fourth cycle. The  $\text{Ag}_3\text{PO}_4$  showed good photocatalytic performance under visible blue light irradiation even after 4 reuse cycles.

The Langmuir-Hinshelwood model (Fig. 2B) [43] was used to determine the photodegradation rate constant  $k$  (angular coefficient). As observed in Fig. 1, the  $k$  constants for the first and fourth cycle are 0.0834 and 0.0801  $\text{min}^{-1}$ , respectively. They are lower because their degradation period is longer. In contrast, the speeds for the second and third cycles are higher, 0.1284 and 0.2223  $\text{min}^{-1}$ , respectively. These results can be attributed to a synergistic effect between the semiconductor and the species generated during the RhB photodegradation process.

The morphological structure of a semiconductor is known to clearly

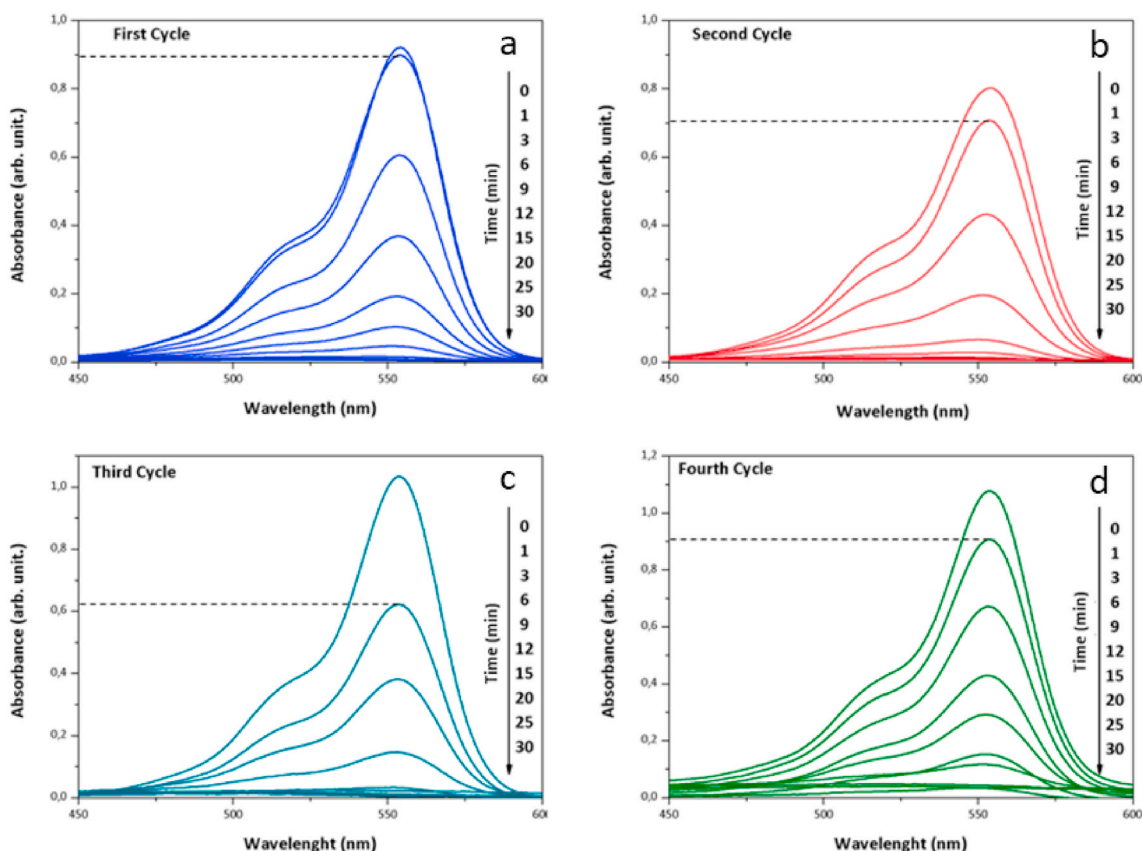


Fig. 1. Photocatalytic activity of  $\text{Ag}_3\text{PO}_4$  after four exposures to visible light.

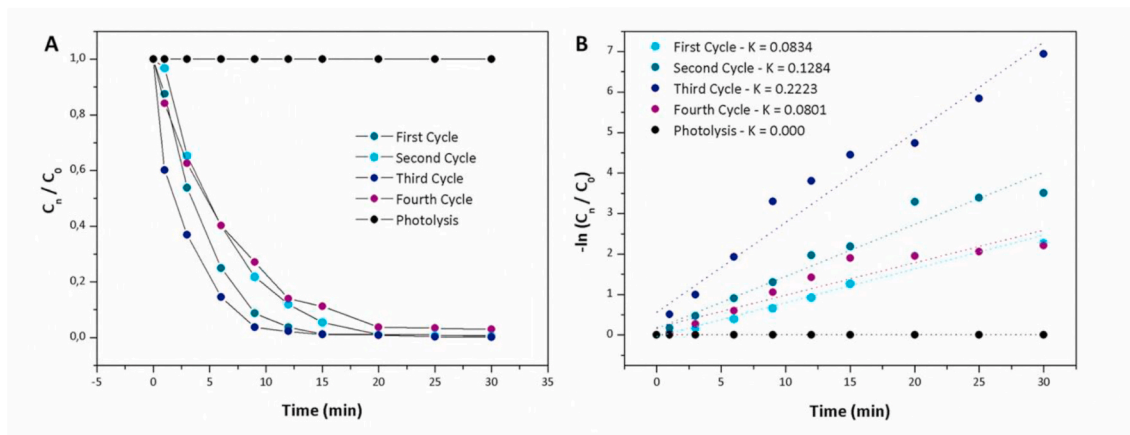
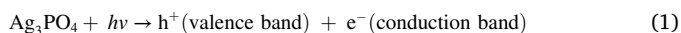


Fig. 2. Rhodamine degradation curve (a) and reaction kinetics (b).

affect photocatalytic performance, as photocatalytic reactions occur on its surface. Previous results of our research group point out that the incomplete local coordination of Ag cations at the exposed surfaces of the  $\text{Ag}_3\text{PO}_4$  semiconductor are the active sites where the surface reactions, that include reduction by electrons,  $e^-$ , and oxidation by holes,  $h^+$ , take place [9].

To propose the mechanism of action of  $\text{Ag}_3\text{PO}_4$  as a photocatalytic, reactive species trapping test was performed. As shown in Fig. S3, when BQ or AO were added to the system, there was a strong inhibition of the photodegradation process. When ISO was added, the photodegradation process occurred without significant changes.

Based on scavenger experiments a possible mechanism for the photocatalytic mechanism of RhB degradation is described as follows:



The generation of reactive oxygen species occurs in the conduction and valence bands of the semiconductor. At the conduction band the  $e^-$  reacts with  $\text{O}_2$  to render  $\cdot\text{O}_2^-$  while  $h^+$  reacts with  $\text{H}_2\text{O}$  to obtain  $\cdot\text{OH}$ . From the results of the scavenger experiments we can assume that the  $\cdot\text{O}_2^-$  radical is responsible for degradation of RhB. The  $\cdot\text{OH}$  radical does not participate directly in this process; however we cannot discard the

presence of hydroxyperoxyl radical ( $\cdot\text{HO}_2$ ). This radical can be obtained by the interaction between  $\cdot\text{O}_2^-$  and  $\text{H}^+$ , formed in the reaction (2). In addition the singlet oxygen,  $^1\text{O}_2$ , can be obtained by means of oxidation:  $\cdot\text{O}_2^- \rightarrow ^1\text{O}_2 + e^-$ .

XRD was performed to verify the order/disorder related to the crystallinity of  $\text{Ag}_3\text{PO}_4$  at a long range. This technique identified parameter changes due to the light cycles, and it determined the phases formed in the pure material and the resulting cycles through the *Inorganic Crystal Structure Database* (ICSD). Fig. 3A shows that the pure material exhibits only the cubic phase of  $\text{Ag}_3\text{PO}_4$ , which is identified by the ICSD 14000 [44] with a cubic structure and belonging to the group  $P43n$  and the main peak (210) located at  $33.2^\circ$ . Moreover, it can be concluded that the material has a good crystallinity due to the sharp definition of the diffraction peaks. As the cycles are performed, cubic and hexagonal metallic Ag are formed. They belong to the space groups  $Fm-3m$  and  $P6_3/mmc$ , respectively, identified by ICSD 44387 [45] and ICSD 56269 [46], respectively. Cubic Ag already appears in the first cycle and becomes more intense until the fourth cycle, where there is more cubic Ag than  $\text{Ag}_3\text{PO}_4$ . The cubic Ag phase is characterized by the appearance of the diffraction planes (111), (200), (220), (311) and (222) located at  $37.9$ ,  $44.5$ ,  $64.4$ ,  $77.3$  and  $81.3^\circ$  respectively. In the third cycle, hexagonal Ag starts to appear, evidenced by the presence of the diffraction planes (110) and (201) located at  $35.9$  and  $40.9^\circ$  respectively. This behavior occurs due to the disassembly of the  $[\text{AgO}_4]$  cluster in  $\text{Ag}_3\text{PO}_4$ , which causes the Ag phase of higher stability (cubic) to be obtained first and become unstable (hexagonal) afterwards [47,48]. Moreover, a centralized amorphization halo was observed at  $24^\circ$ . They

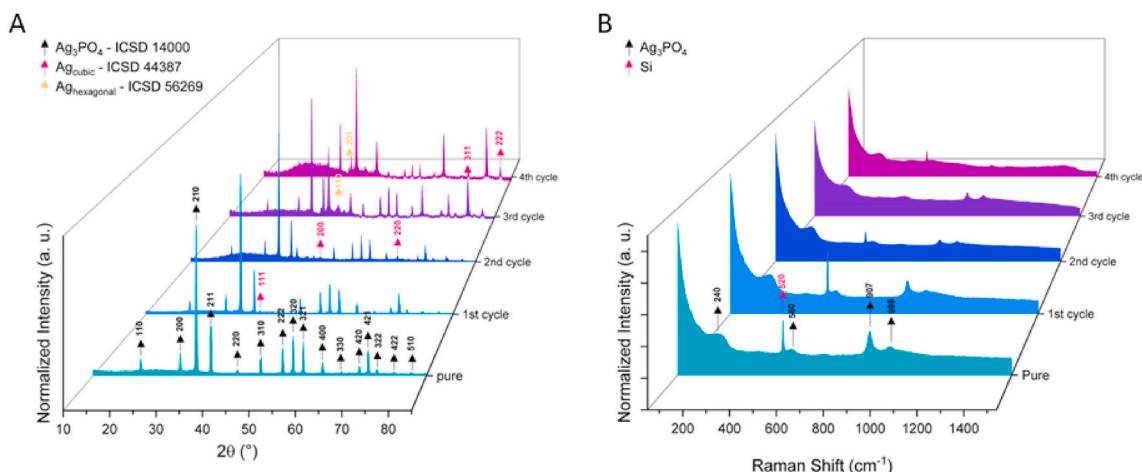


Fig. 3. (A) XRD patterns of  $\text{Ag}_3\text{PO}_4$  after four exposures to visible light and (B) Micro-Raman spectroscopy of  $\text{Ag}_3\text{PO}_4$  after four exposures to visible light.



increase proportionally to the number of cycles, demonstrating the low crystallinity of the system. This phenomenon occurs through the irradiation of the material, where in the  $[\text{AgO}_4]$  clusters the reduction process takes place, leading to the formation of metallic Ag. This process favors the decomposition of  $\text{Ag}_3\text{PO}_4$ , which leads to the formation of an amorphous material composed of  $\text{Ag}_x\text{P}_y\text{O}_z$  and reduces the crystallinity of the final system.

When the formation of cubic Ag occurs during the photodegradation process, a heterostructure is formed,  $\text{Ag}_{\text{cubic}}/\text{Ag}_3\text{PO}_4$ . The synergistic effect between these two phases increases the RhB photodegradation rate in the second and third cycles, as shown in Figs. 1 and 2, respectively. In contact with electromagnetic radiation, the electrons of the Ag nanoparticles (Ag NPs) provoke the surface plasmon resonance effect. These plasmon oscillations promote the electronic transfer processes, increasing the efficiency of heterostructures for photocatalytic processes [49]. In the fourth cycle, the photocorrosion process causes hexagonal Ag to be formed together with  $\text{Ag}_{\text{cubic}}/\text{Ag}_3\text{PO}_4$ , also reducing the rate of degradation. Thus, the formation of this specific phase increases the electron-hole recombination ( $e^-h^*$ ) of the system, which makes the process less efficient than the previous ones.

Micro-Raman spectroscopy was performed to analyze order/disorder of the system at short range (Fig. 3B). According to the group theory, through the decomposition of the  $\Gamma$  point,  $\text{Ag}_3\text{PO}_4$  has 18 active modes in the Raman, given by  $2A_1 + 4E + 12T_2$ . However, due to the high disorder at short range, only four modes were observed, at 240, 560, 907, and  $998\text{ cm}^{-1}$ . The  $240\text{ cm}^{-1}$  ( $T_2$ ) mode is related to the characteristic rotation/translation of the  $[\text{PO}_4]$  cluster. The mode at  $560\text{ cm}^{-1}$  (E) refers to the asymmetric bending of the  $[\text{PO}_4]$  cluster. The modes at 907 ( $A_1$ ) and  $998\text{ cm}^{-1}$  ( $T_2$ ) refer to symmetrical and asymmetrical stretching, respectively, and the symmetrical stretching is more intense. As the material is recycled through photocatalysis, an increase of the short-range disorder occurs due to the dissipation of the modes mentioned above. This behavior can be attributed to the structural degradation of  $\text{Ag}_3\text{PO}_4$  during the photocatalytic cycles, which favors the formation of secondary materials (cubic and hexagonal Ag, observed in the XRD). Thus, the characteristic modes of the matrix become less intense, due to the increase in the structural disorder of  $\text{Ag}_3\text{PO}_4$ .

Fig. 4 shows the FE-SEM micrographs for the pure  $\text{Ag}_3\text{PO}_4$  and after the RhB photodegradation cycles. The characteristics of the photocatalysis depend on the morphology of the material. Based on the parameters used in the synthesis, such as temperature and surfactants, several morphologies can be obtained such as polyhedra, circles, and particles with irregular morphology. In this study, the CP method applied to the synthesis of  $\text{Ag}_3\text{PO}_4$  presented a morphology of agglomerated irregular spheres (Fig. 4A and B). As the RhB photodegradation cycles were performed, degradation on the surfaces of these spheres was observed, as shown in Fig. 4C–J. This occurred due to the degradation of  $\text{Ag}_3\text{PO}_4$  for the formation of metallic Ag, which was confirmed by XRD

and Micro-Raman spectroscopy measurements. It was also observed that particle size decreases with increasing photocatalytic cycles, ranging from  $294 \pm 68\text{ nm}$  for the pure material to  $107 \pm 32$  for the material obtained in the last cycle (for the first, second, and third cycles,  $250 \pm 51$ ,  $217 \pm 53$ , and  $142 \pm 47\text{ nm}$  sizes were obtained, respectively). These results can be attributed to the degradation of  $\text{Ag}_3\text{PO}_4$  in the photocatalytic process, which leads to the formation of other species, such as cubic and hexagonal Ag.

## 2.2. Antifungal activity of $\text{Ag}_3\text{PO}_4$ in the presence and absence of light

The antifungal activity of  $\text{Ag}_3\text{PO}_4$  in different concentrations was evaluated against *C. albicans* ATCC 90028 in planktonic and biofilm in formation, in the presence (L+) and absence (L-) of visible-light irradiation (455 nm). A difference in inhibitory concentrations (IC) was observed between L+ and L-groups.

Two-way ANOVA showed that there is an interaction ( $p < 0.001$ ) between the factors analyzed (light and concentration) on *C. albicans*, both in suspension and in biofilm formation. In *C. albicans* in suspension, two-way ANOVA showed that there is an effect of light [ $F(1; 208) = 62.742$ ;  $p < 0.001$ ], concentration [ $F(12; 208) = 345.651$ ;  $p < 0.001$ ], as well as the interaction between light and concentration [ $F(12; 208) = 44.528$ ;  $p < 0.001$ ] on the viability of *C. albicans*.

The suspension data was analyzed starting at  $2000\text{ }\mu\text{g/mL}$  and decreasing to  $0\text{ }\mu\text{g/mL}$  (control). Considering whether or not to use light at the same concentration of  $125$  or  $62.5\text{ }\mu\text{g/mL}$ , the mean  $\text{Log}_{10}$  (CFU/mL) in the L+ were significantly higher than those in the L-. The results in the L- showed that CFU started to decrease at  $62.5\text{ }\mu\text{g/mL}$  and decreased at higher concentrations of  $\text{Ag}_3\text{PO}_4$ . In L+, the mean CFU was significantly lower at  $125\text{ }\mu\text{g/mL}$  compared to the remaining minor concentrations ( $\leq 23\%$  of reduction). Although presenting a higher percentage of kill in sub-MFC in L- (concentration of  $1000\text{ }\mu\text{g/mL}$ , with a reduction of 42%) compared to the sub-MFC in L+ (concentration of  $125\text{ }\mu\text{g/mL}$ , with a reduction of 23%), there was a significant reduction in particle concentration required to completely eliminate microorganisms from  $2000$  to  $250\text{ }\mu\text{g/mL}$ , suggesting the efficacy of a lower silver phosphate particle concentration for MFC in L+.

The inhibition of microorganism was total whenever light was present at  $250\text{ }\mu\text{g/mL}$  or higher concentrations. However, in the absence of light, inhibition was only complete at the highest concentration of  $\text{Ag}_3\text{PO}_4$  ( $2000\text{ }\mu\text{g/mL}$ ).

In biofilm formation, two-way ANOVA also showed that there is an effect of visible light [ $F(1; 96) = 43.121$ ;  $p < 0.001$ ], and concentration [ $F(5; 96) = 141.428$ ;  $p < 0.001$ ], as well as the interaction between light and concentration [ $F(5; 96) = 28.374$ ;  $p < 0.001$ ] on the viability of *C. albicans* cells. The absence and presence of visible light and the microcrystal concentrations ( $0\text{ }\mu\text{g/mL}$  to  $1000\text{ }\mu\text{g/mL}$ ) were used as fixed factors. At  $1000\text{ }\mu\text{g/mL}$ , the mean  $\text{Log}_{10}$  (CFU/mL) was

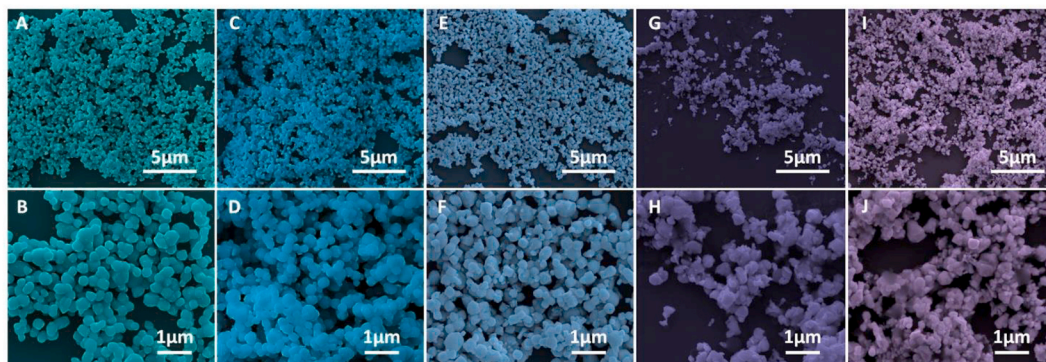
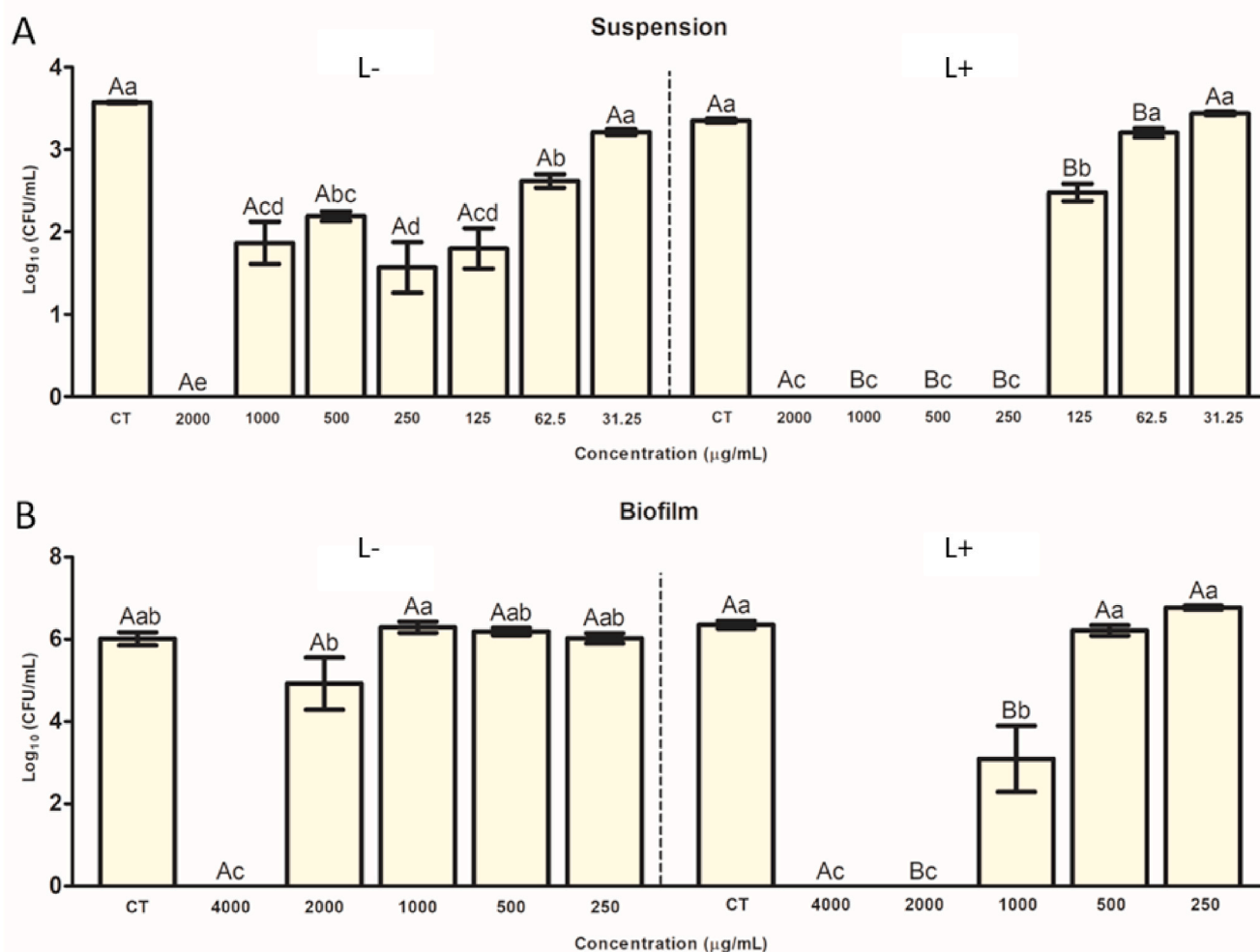


Fig. 4. FE-SEM of  $\text{Ag}_3\text{PO}_4$  microcrystals on different magnifications. Pure (A), after one (C), two (E), three (G), and four (I) exposures to visible-light and the respective magnifications at B, D, F, H, and J.





**Fig. 5.** Mean values of Log<sub>10</sub>(CFU/mL) of *C. albicans* in suspension (A) and biofilm formation (B) after treatment with different concentrations of Ag<sub>3</sub>PO<sub>4</sub> in the presence and absent visible-light irradiation. CT: control; L-: without visible-light irradiation; L+: presence of visible-light irradiation. Error bars: standard deviation. Equal lowercase letters denote equality of means within the group (L+ or L-); Equal uppercase letters denote equality of means in the same concentration between the groups.

significantly lower in L+ (reduction of 23%) than in L- (no reduction), and it was equivalent in other concentrations (Fig. 5). In L+, the CFU averages were equivalent at concentrations from 500 to 0 µg/mL. The mean was lower for 1000 µg/mL, and higher and equivalent for other concentrations.

The data presented by the biofilm showed the minimal biofilm inhibitory concentration (MBIC) at 4000 µg/mL (100% of reduction) in L-, and at 2000 µg/mL in L+. Both sub-MBIC, in L- (2000 µg/mL) and L+ (1000 µg/mL), showed reduction compared to the control (6,5 % and 23 %, respectively), but the presence of light suggested a greater efficacy using lower silver phosphate particle concentration.

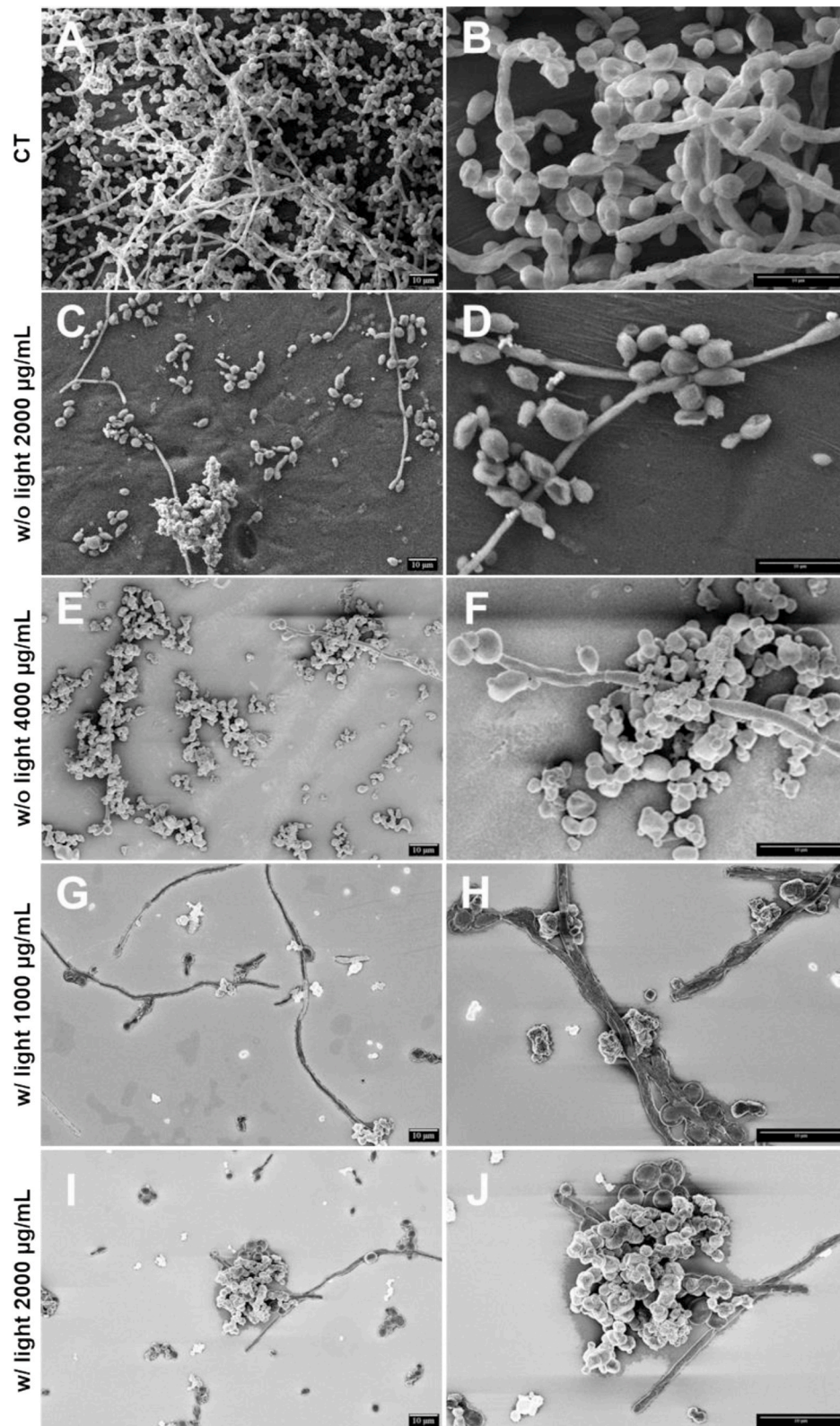
This information is in agreement with FE-SEM images (Fig. 6). A reduction of *C. albicans* was observed when compared to the image control microorganisms without Ag<sub>3</sub>PO<sub>4</sub>. In the FE-SEM images (Fig. 6), we can also observe a reduction in the growth of *C. albicans*, however, with no elimination of fungus. Nevertheless, the morphology of the blastopores was modified from round to irregular (Fig. 6D and H).

An analysis of confocal microscopy (Fig. 7) shows that the red staining (propidium iodide) becomes more intense as the concentrations increase, and it is inversely proportional to the green staining (SYTO 9). Moreover, the control group demonstrated a cluster of living microorganisms, which progressively decreased in other concentrations. These other concentrations presented black spaces, which may be characteristic of microorganism reduction. These results are in accordance with

the Log<sub>10</sub> (CFU/mL) data, where the difference between the control group and IC is significant ( $p < 0.001$ ).

The microcrystals agglutinated throughout the tests to evaluate the inhibitory activity of the Ag<sub>3</sub>PO<sub>4</sub> microcrystals against *C. albicans* during its biofilm formation. Methodological optimizations were necessary, changing the culture medium from RPMI to YNB, to avoid this problem and to allow an appropriate distribution of concentrations. The literature is clear regarding the growth of microorganisms according to the culture medium used, that is, the culture medium affects the growth, adhesion, and development of biofilms [50]. According to the previously mentioned study, the growth of *C. albicans* biofilm is favored when the RPMI medium is used, supporting the growth of hyphae and extracellular matrix [51], which are characteristics of a well-established biofilm. In this study, we chose to perform the tests with the YNB medium since the material agglutinated when in contact with the RPMI medium. The use of the YNB culture medium can reduce the metabolic activity of *C. albicans* biofilm, resulting in a biofilm with less evident characteristics [51], such as fewer hyphae development. These characteristics can be seen in Fig. 7.

As the particles are irradiated, morphological changes take place on the surface of the material. These changes are responsible for the release of Ag ions, which can interact with the plasma membrane, DNA, and enzymatic processes of the microorganisms, leading to their death [52, 53]. In addition, the energy provided by the photons and the surface



**Fig. 6.** FE-SEM of *C. albicans* biofilm formed in the presence of  $\text{Ag}_3\text{PO}_4$  microcrystals without (w/o) and with visible-light irradiation (w/). The images (A) and (B) are control; (C–D) and (E–F) are 2000  $\mu\text{g/mL}$  and 4000  $\mu\text{g/mL}$ ; (G–H) and (I–J), 1000  $\mu\text{g/mL}$  and 2000  $\mu\text{g/mL}$ . The (B), (D), (F), (H), (H) images are magnifications of (A), (C), (E), (G) and (I) images.



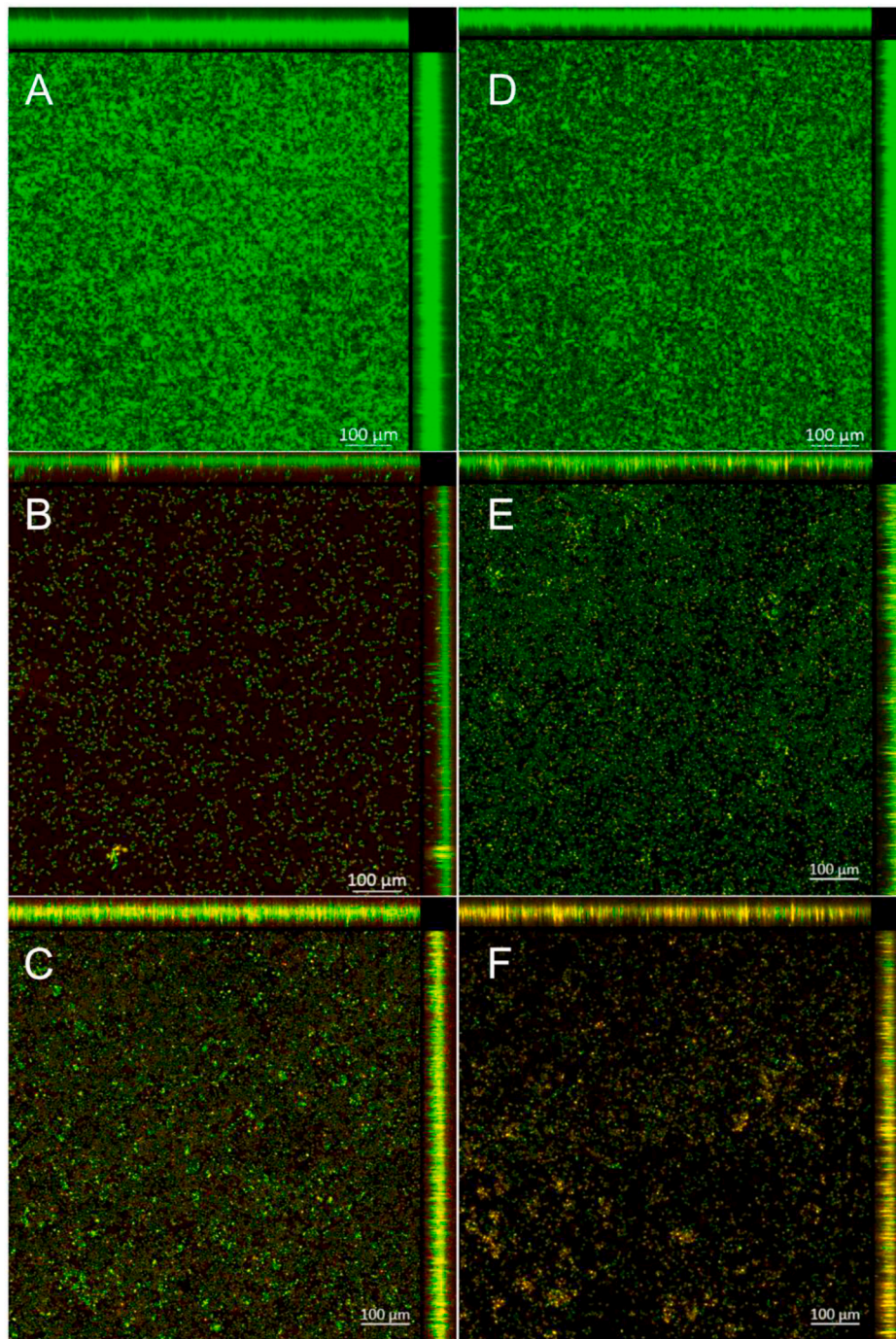


Fig. 7. Confocal microscopy of *C. albicans* biofilm formed in the presence of  $\text{Ag}_3\text{PO}_4$  microcrystals with (A, B, C) and without visible-light irradiation (D, E, F). A and D represent the control group, followed by B and C (2000 and 4000  $\mu\text{g}/\text{mL}$ ), and E and F (1000 and 2000  $\mu\text{g}/\text{mL}$ ), with  $\text{Ag}_3\text{PO}_4$  concentrations used against the biofilm.

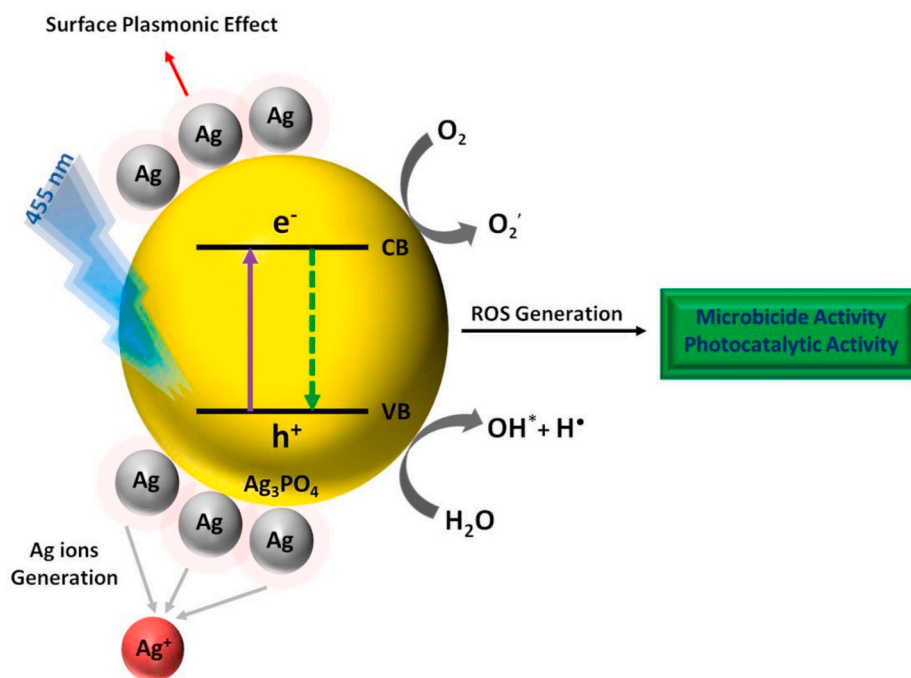
plasmon resonance effect facilitates the migration of electrons. The main factor responsible for both RhB degradation and antifungal activity under visible-light irradiation is the same. It comes into contact with the cell wall and membrane, breaking them both. It also generates oxidative stress inside the cell, disrupting its intracellular structures, which can lead to cell death [54–56]. Moreover, ROS is generated inside the cells, possibly due to silver ions which inhibit the respiratory enzyme, which attacks itself [57]. Fig. 8 shows the schematic drawing of the mechanism of action of the  $\text{Ag}_3\text{PO}_4/\text{Ag}$  heterostructure when irradiated with blue light (455 nm).

Investigations were carried out to evaluate the growth rate of

microorganisms in the presence of  $\text{Ag}_3\text{PO}_4$  [58]. However, there are no studies in the literature regarding their action both against biofilm and in the presence of the fungus *C. albicans*, as analyzed in the present study. Compared to the suspension, the biofilm presented higher inhibitory and subinhibitory concentrations due to its greater complexity and the presence of an extracellular matrix, which is an inherent resistance of biofilms [59].

When the electrons of  $\text{Ag}_3\text{PO}_4$  are excited from VB to CB,  $\text{Ag}_3\text{PO}_4$  produces ROS and releases silver ions due to its gradual degradation [60]. These results corroborate with the data provided by the XRD, micro-Raman, and FE-SEM measurements, which show changes in





**Fig. 8.** A schematic representation of the mechanism for photodegradation and microbicidal activity of synthesized  $\text{Ag}_3\text{PO}_4$  microcrystals submitted to irradiation with visible-light (455 nm). SPE: surface plasmonic effect;  $e^-$ : electron;  $h^+$ : hole; CB: conduction band; VB: valence band.

$\text{Ag}_3\text{PO}_4$  at long and short ranges. The structural degradation of  $\text{Ag}_3\text{PO}_4$  for the formation of metallic Ag was directly proportional to the morphological alteration of the material. Thus, it is suggested that a certain degree of disorder, both structural and morphological, is desirable for greater system efficiency.

### 3. Experimental

#### 3.1. Synthesis

$\text{Ag}_3\text{PO}_4$  microcrystals were synthesized at room temperature by the CP method because it is a simple, low cost and fast method that does not require heat treatments, besides being widely used in the literature [61–63]. For that,  $3 \times 10^{-3}$  mol of silver nitrate ( $\text{AgNO}_3$ , Cennabras, 98%) and  $1 \times 10^{-3}$  mol of diammonium hydrogen phosphate ( $[\text{NH}_4]_2\text{HPO}_4$ , Alfa-Aesar, 98%) were separately dissolved in beakers containing 75 and 25 mL of distilled water, respectively. After complete dissolution, the precursors were mixed for 10 min under magnetic stirring, promoting the instant precipitation of  $\text{Ag}_3\text{PO}_4$ . The resulting precipitate was washed several times with distilled water and acetone to remove remaining ions in contact with the material. Subsequently, the obtained  $\text{Ag}_3\text{PO}_4$  was centrifuged and oven dried at 60 °C.

#### 3.2. Characterization

The  $\text{Ag}_3\text{PO}_4$  microcrystals were characterized by XRD using a Rigaku D/Max 2500 PC (Japan) diffractometer. A scanning angle of 10 to 85°, scanning speed of 1°/min, angular pitch of 0.02°, and  $\text{CuK}\alpha$  radiation ( $\lambda = 1.5406 \text{ \AA}$ ) were used. To determine the band gap, a Varian Cary 5G spectrometer was used at a wavelength of 200 to 800 nm and scanning speed of 600 nm/min, using the Kubelka-Munk function [64,65]. The photocatalytic activity was evaluated considering the degradation of rhodamine B (RhB) in an aqueous solution under visible-light irradiation [1]. Thus, 50 mg of  $\text{Ag}_3\text{PO}_4$  powder was mixed in 50 mL of  $1 \times 10^{-5}$  mol/L RhB solution for 15 min in an ultrasonic bath (1510 Branson, 42 kHz). The system was magnetically stirred in the dark for 30 min at 20 °C to promote adsorption-desorption between the dye and the powder.

The temperature was controlled by a thermostatic bath. When the first aliquot of 2 mL was collected, the time was denominated as 0 min. Subsequently, there was an optical system, called “biotable”, composed of 24 LEDs with 455-nm wavelength and 31  $\text{mW}/\text{cm}^2$  irradiance power in the suspension. In this system, aliquots were collected at different times: 0 (control, without the presence of  $\text{Ag}_3\text{PO}_4$ ), 1 (1.86  $\text{J}/\text{cm}^2$ ), 3 (5.58  $\text{J}/\text{cm}^2$ ), 6 (11.16  $\text{J}/\text{cm}^2$ ), 9 (16.74  $\text{J}/\text{cm}^2$ ), 12 (22.32  $\text{J}/\text{cm}^2$ ), 15 (27.9  $\text{J}/\text{cm}^2$ ), 20 (37.2  $\text{J}/\text{cm}^2$ ), 25 (46.5  $\text{J}/\text{cm}^2$ ), and 30 (55.8  $\text{J}/\text{cm}^2$ ) minutes. The aliquots were centrifuged (MiniSpin) for 3 min (8300 rpm) and analyzed in the UV–Vis spectrophotometer to monitor the maximum absorption band of RhB ( $\lambda = 554 \text{ nm}$ ). To analyze the order/disorder of the system at short-range and the morphology and size of the samples, micro-Raman spectroscopy and FE-SEM (Carl Zeiss Microscope operated at 30 kV) were performed, respectively.

In order to propose a mechanism for the photocatalytic and fungicidal activity of  $\text{Ag}_3\text{PO}_4$ , reactive species trapping test was performed as described previously [41,66]. Benzoquinone (BQ), ammonium oxalate (AO) and isopropanol (ISO) as scavengers for superoxide radical ( $\cdot\text{O}_2^-$ ), holes ( $h^+$ ) and hydroxyl radical ( $\cdot\text{OH}$ ), respectively.

#### 3.3. Antifungal tests using synthesized $\text{Ag}_3\text{PO}_4$

The synthesized  $\text{Ag}_3\text{PO}_4$  microcrystals were tested to verify their antifungal ability against a standard strain of *C. albicans* from the American Type Culture Collection (ATCC) 90028. The test determined their MFC and MBIC [67] in both, planktonic and biofilms cells, in the presence and absence of light.

The frozen microorganisms were reactivated in a Sabouraud Dextrose Agar (SDA - Michigan, USA) culture medium with 5  $\mu\text{g}/\text{mL}$  chloramphenicol, and incubated at 37 °C for 48 h. After this period, 5 colonies of similar size were transferred into 5 mL of Yeast Nitrogen Base (YNB - Maharashtra, India) specific culture medium in a Falcon tube, which was incubated at 37 °C for 16 h. Subsequently, 500  $\mu\text{L}$  of pre-inoculum were diluted in 9.5 mL of YNB and incubated again for 8 h at 37 °C. The growth phase was determined by the optical density (OD 540 nm) of the suspension, which was standardized in a spectrophotometer at 0.55 ( $\pm 0.08$ ), with a concentration of approximately  $7.94 \times$

$10^6$  CFU/mL.

### 3.3.1. Microbiological tests in planktonic cells

After OD was reached, the growth rate was diluted ( $10^6$  to  $10^3$ ). Subsequently, microcrystals diluted in 2 mL microtubes, with concentrations between 0.9 and 2000  $\mu\text{g/mL}$ , were placed in 24-well plates (Kasvi–China). Samples of *C. albicans* at  $10^3$  were added to each well of the 24-well plates, for a total of 500  $\mu\text{L}$  (250  $\mu\text{L}$  of microcrystal and 250  $\mu\text{L}$  of *C. albicans* suspension). Two plates were used: one exposed to visible-light (L+) and the other preserved in the dark (L-). Shortly after the contact between microorganisms and microcrystals, one plate was wrapped in aluminum foil and remained incubated at 37 °C for 30 min. The other plate was exposed to light at a 455-nm wavelength (55.8 J/cm<sup>2</sup>) for the same period. After that, both plates were plated in duplicate by the drip method (10  $\mu\text{L}$ ) in Petri dishes containing SDA culture medium with chloramphenicol. The dishes were maintained in a static incubator at 37 °C for 24 h for subsequent counting of colony forming units (CFU/mL). The analyses were performed in triplicate and on three different occasions.

### 3.3.2. Microbiological tests of biofilms in formation

After OD was reached, *C. albicans* was added to two 24-well plates (TPP–Trasadingen, Switzerland) at a concentration of  $10^7$  (2 mL). The plates were placed in an orbital shaker (37 °C, 75 rpm) for 90 min to provide initial adhesion [68]. Subsequently, each well was washed two times with phosphate-buffered saline (PBS, 1 mL) to remove the weakly adhered microorganisms. After that, the wells were resuspended with 1 mL of YNB medium and 1 mL of microcrystals diluted in PBS, for a total of 2 mL with different concentrations in each well [69]. One plate was wrapped in aluminum foil and remained in the orbital shaker at 37 °C, while the other was exposed to visible-light at a 455-nm wavelength for 30 min. A blue (455 nm) light-emitting diode was chosen due to its great use in photoinactivation of oral microorganisms like *C. albicans* [70,71]. Next, the plate exposed to light was also wrapped in aluminum foil, and both plates remained in the orbital shaker at 37 °C for 48 h. Subsequently, both plates were removed from the orbital shaker, washed with 1 mL of PBS (three times), and resuspended in 2 mL of YNB. To count the colony forming units (CFU/mL), the biofilms were removed from the bottom of the plate with the aid of a sterile tip, and 20  $\mu\text{L}$  aliquots of the detached biofilms were serially diluted in PBS ( $10^{-1}$  to  $10^{-4}$ ). Afterwards, 10  $\mu\text{L}$  aliquots of each dilution ( $10^0$  to  $10^{-4}$ ) were seeded in dishes containing SDA by the drip method (10  $\mu\text{L}$ ). The sowing procedures were performed in duplicate. Then, the Petri dishes were incubated for 24 h at 37 °C for subsequent counting of the colony forming units (CFU/mL). The LED device used, covered the wavelength range from 440 to 460 nm, with maximum emission at 455 nm and the analyses were performed in triplicate and on three different occasions.

### 3.3.3. Microscopy analyses of biofilm with microcrystals

The biofilm was formed in two 24-well plates ( $n = 2$ ), and the growth protocol was followed as described above, resulting in a concentration of approximately  $1 \times 10^7$  CFU/mL. After a 48h incubation, the particles were washed (three times), and the biofilm was marked with the fluorophores SYTO 9 and propidium iodide, which were present in the LIVE/DEAD kit and handled according to the manufacturer's instructions. The biofilms were maintained in PBS for visualization on the confocal fluorescence microscope (Carl Zeiss LSM 780, Thuringia, Germany) with 488 nm (excitation) and 561 nm (emission) lasers on the 20x objective. The images were analyzed in the ZEN Blue software version 2.3 (Thuringia, Germany) and this experiment was performed in triplicate in two independent occasions.

To analyze the morphology of the biofilm and microcrystals, FE-SEM (Thuringia, Germany) was used. For the biofilm analysis, polystyrene discs were cut from the bottom of a 24-well plate (Trasadingen, Switzerland). The discs were used with the biofilm during its growth in the bottom of the plate, following the same growth conditions described

above, and using a concentration of approximately  $1 \times 10^7$  CFU/mL. To sterilize the discs, they were added to a beaker containing 200 mL of distilled water and placed in a 645-W microwave for 3 min [72].

After 48 h of incubation, each well was washed with PBS (three times), and the disks that remained in the bottom throughout the biofilm growth were immersed for 20 min in paraformaldehyde and serially dehydrated in ethanol (70%, 80%, 90%, and 100%) for 20 min at each concentration. The disks were dried at room temperature and kept in a vacuum desiccator until the analysis, for which the field emission scanning electron microscope (Thuringia, Germany) was used. The obtained images used magnifications of 500x and 2500x at 10 kV and wavelength dispersion (WDS) of 8 mm. Before microscopic observation, the samples were mounted on metallic stubs and covered with gold.

### 3.4. Statistical analysis

In the present study, the concentration and the presence or absence of light were considered to be a variation factor. After verifying the normality and homogeneity of data, ANOVA two-way test was applied followed by posthoc Tukey ( $\alpha = 0.05$ ) was performed. The statistical analysis was carried out using the software SPSS (IBM, NY, USA).

## 4. Conclusions

The main conclusions of the present work can be summarized as follows: (i)  $\text{Ag}_3\text{PO}_4$  microcrystals were synthesized by the CP method, and after four cycles of light exposure at a 455-nm wavelength, cubic (from 1 exposure) and hexagonal  $\text{Ag}$  (from 3 exposures) were formed; (ii) after each exposure to light, it was observed that the  $[\text{AgO}_4]$  clusters, as building blocks of the 3D structure of  $\text{Ag}_3\text{PO}_4$ , are the sites where the formation of metallic  $\text{Ag}^0$  takes place. This process favors material decomposition by altering its morphology at the end of the fourth light exposition, (iii) in the microbiological tests with and without light,  $\text{Ag}_3\text{PO}_4$  presented antifungal activity for both planktonic and biofilm forms. However, it was observed that its antifungal activity was more effective with lower concentrations in the presence of light when compared to higher microcrystal concentrations in the absence of light; (iv) the antifungal activity of  $\text{Ag}_3\text{PO}_4$  was increased by visible-light irradiation; and (v) based on the present results related to antimicrobial capacity, durability and long-term use, we believe this study is of fundamental importance to the design of photocatalytic and fungicide  $\text{Ag}_3\text{PO}_4$  based materials.

### Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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### Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.ceramint.2021.04.272>.

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